

Evaluations of *Bacillus* species against *Callosobruchus chinensis*, *Callosobruchus maculatus* (Coleoptera: Tenebrionidae) under laboratory and store conditions

Abstract: Sabbour Magda¹ and Maysa E. Moharam²

1. Pests and Plant Prot., Dept . National Research Centre, Cairo Egypt
El-Tahrir St. Dokki, Cairo, Egypt

sabbourm@yahoo.com or magasabbour@gmail.com

2. Microbial Chemistry Department, Genetic Engineering & Biotechnology
Division.e. mail. maysa_nrc@live.com

Abstract: Five strains of the entopathogenic bacteria *Bacillus thuringiensis* were evaluated against two stored products insect pests, *Callosobruchus chinensis*, *Callosobruchus maculatus* (Coleoptera: Tenebrionidae). The LC50s of *C. maculatus* of different tested bacteria which recorded that, 248, 269, 144, 88 and 110 Ug/ml after *C. maculatus* treated with different concentrations of *B.T J*, *B.t 0900*, *Bt NRRL 2172*, *BT IP thurizide* and *Bt HD112*., under laboratory conditions ., respectively. The corresponding LC50s of *C. chinensis*, 233, 132, 145, 77 and 100 Ug/ml ., respectively .

Under store conditions the number of eggs laid per female of *C. maculatus* were significantly decreased and recorded 1.7 ± 2.7 , 5.3 ± 0.5 , 10.2 ± 2.3 , and 11.4 ± 3.7 eggs/female after 20, 45, 90 and 120 days of storage after treated with *BT IP thurizide* ., respectively as compared 1.7 ± 2.7 to 19.8 ± 2.5 , 89.3 ± 4.5 , 91.5 ± 5.5 and 99.8 ± 1.9 eggs/female in the control. Under store conditions, the percentage of eggs laid/female were significantly decreased to 11.0 ± 1.7 eggs/female after *C. chinensis* treated with *BT IP thurizide* after 120 days as compared to 98.9 ± 1.9 eggs/female in the control. In all bacterial treatments the percentage of emergence were significantly decreased to 1, 1, 1, 4 and 6 with treated with *BT IP thurizide* .

Keywords: *Bacillus* species, *Bacillus* species, *B.T J*; *B.t 0900*; *Bt NRRL 2172*; *BT IP thurizide*; *Bt HD112*; *Callosobruchus chinensis* *Callosobruchus maculatus* (Coleoptera: Tenebrionidae)

1. Introduction

The common bean (*Phaseolus vulgaris* L.) is one of the principal food and cash crop legume grown in the tropical world [11]; [2] ; [3]. Almost all the insect

pests of stored grains have a remarkably high rate of multiplication and within one season, they may destroy 10- 15% of the grains and contaminate the rest with undesirable odours and flavours [4]. A wide range of seed beetles attack the grain of common bean varieties [5]. However, the predominant damaging pests of stored grain legumes mainly in the tropics are *Callosobruchus maculatus* (Fab.), *C. chinensis* (L.), *Caryedon serratus* (Oliver), *Zabrotes subfasciatus* (Boheman) and *Acanthoselids obtectus* (Say) ([6]; [7]; [8]).

Essential oils may have attractive or repellent effects and in some cases they showed an insecticidal action against insects. Essential oils isolated from plants and consisting of cyclic and monocyclic mono-terpenes are effective repellents against insects ([9],[10]). Oil carriers can also distribute the inoculum over the thin intersegmental membranes, which are more readily penetrated by entomopathogenous fungi[6].

The present work aimed to explore the protective potency of some microbial bacteria *Bacillus thuringiensis* against, broad bean beetle, *C. maculatus* and *C. chinensis* during storage.

Sabbour, 2012, 2014, Sabbour and Abd El-Aziz, 2007a, 2007b, 2010;

2. Materials and Methods

2.1. Tested Insects:

The broad bean beetle (cowpea beetle), *Callosobruchus maculatus* (F.) and *C. chinensis* (L.) were reared on broad bean seeds *Vigna faba* (L.) at 28±2°C and 60±5% R.H. under laboratory conditions.

2.2. Microorganisms:

Bacillus thuringiensis 09001, *Bacillus thuringiensis* NRRL 2172, *Bacillus thuringiensis* IP thuricide, *Bacillus thuringiensis* HD 112, and *Bacillus thuringiensis* J were used in this study. The bacterial cultures were maintained on nutrient agar slants at 4°C.

2.3. Bacterial culture media:

The conventional laboratory culture broth, Nutrient broth , was used for culture preparation by mixing 5g peptone and 3g beef extract/ 1 L dist water. 50 ml of sterile

medium was inoculated with one loopful of bacterial strain and incubated under shaking growth conditions on an orbital rotary shaker (125rpm) at 30°C for 72h.

2.4. Effect of the Microbial Control Agents: Isolated *Bacillus thuringiensis* (Bt) B.T J; *B.t* 0900; *Bt* NRRL 2172; *BT* IP thurizide; *Bt* HD112; were used to test their activities on stored insect pests *Sitophilus oryzae* adult beetles. The dead larvae of *S. oryzae* were collected from the colony. The pathogen were isolated according to Salama et al [24]. The of Bt the tested concentrations were (500, 250, 125, 63, 32 and 16 ug/ml) (w/v). The rice pots were sprayed by tested concentrations of fungi or Bt and left to dry under laboratory conditions. Control treatment was made by feeding the larvae on untreated rice. The percentages of mortality were counted and calculated according to 50 [17], while LC50 were calculated through probit analysis according to [18]. The experiments were carried under laboratory conditions; 26 ± 2o C and 60- 70% R.H.

2.5. Effect of Storage Period on Weight Loss: To determine the impact of storage period on weight loss in the studied cultivars, samples of seeds were tested and as previously mentioned above during storage and weight loss was calculated according to

Harris and Lindblad :

$$\text{Weight loss \%} = \frac{(w_u \times n_d) - (w_d \times n_u)}{W_u (n_d + n_u)} \times 10$$

Where:

Wd= weight of damaged seeds

nu= number of undamaged seeds

wu= weight of undamaged seeds

nd= number of damaged seeds

Data were subjected to analysis of variance (ANOVA) and means were compared by a least significant different test.

3. Results

2.2. Microorganisms:

Bacillus thuringiensis 09001, *Bacillus thuringiensis* NRRL 2172, *Bacillus thuringiensis* IP thuricide, *Bacillus thuringiensis* HD 112, and *Bacillus thuringiensis* J were used in this study. The bacterial cultures were maintained on nutrient agar slants at 4°C.

2.3. Bacterial culture media:

The conventional laboratory culture broth, Nutrient broth, was used for culture preparation by mixing 5g peptone and 3g beef extract/ 1 L dist water. 50 ml of sterile medium was inoculated with one loopful of bacterial strain and incubated under shaking growth conditions on an orbital rotary shaker (125rpm) at 30°C for 72h.

2.4. Effect of the Microbial Control Agents: Isolated *Bacillus thuringiensis* (Bt) B.T J; *B.t 0900*; *Bt NRRL 2172*; *BT IP thurizide*; *Bt HD112*; were used to test their activities on stored insect pests *Sitophilus oryzae* adult beetles. The dead larvae of *S. oryzae* were collected from the colony. The pathogen were isolated according to Salama et al [24]. The of Bt the tested concentrations were (500, 250, 125, 63, 32 and 16 ug/ml) (w/v). The rice pots were sprayed by tested concentrations of fungi or Bt and left to dry under laboratory conditions. Control treatment was made by feeding the larvae on untreated rice. The percentages of mortality were counted and calculated according to 50 [17], while LC50 were calculated through probit analysis according to [18]. The experiments were carried under laboratory conditions; 26 ± 2o C and 60- 70% R.H.

2.5. Effect of Storage Period on Weight Loss: To determine the impact of storage period on weight loss in the studied cultivars, samples of seeds were tested and as previously mentioned above during storage and weight loss was calculated according to

Harris and Lindblad :

$$\text{Weight loss \%} = \frac{(w_u \times n_d) - (w_d \times n_u)}{W_u (n_d + n_u)} \times 10$$

Where:

Wd= weight of damaged seeds

nu= number of undamaged seeds

wu= weight of undamaged seeds

nd= number of damaged seeds

Data were subjected to analysis of variance (ANOVA) and means were compared by a least significant different test.

2. Results

Table 1 show that the LC50s of *C. maculatus* of different tested bacteria which recorded that, 248, 269, 144, 88 and 110 Ug/ml after *C. maculatus* treated with different concentrations of *B.T J*, *B.t 0900*, *Bt NRRL 2172*, *BT IP thurizide* and *Bt HD112*., under laboratory conditions ., respectively (Table1).

The corresponding LC₅₀s of *C. chinensis*, 233, 132, 145, 77 and 100 U_g/ml., respectively (Table 2).

Under store conditions the number of eggs laid per female of *C. maculatus* were significantly decreased and recorded 1.7 ± 2.7 , 5.3 ± 0.5 , 10.2 ± 2.3 , and 11.4 ± 3.7 eggs/female after 20, 45, 90 and 120 days of storage after treated with *BT IP thurizide*., respectively as compared 1.7 ± 2.7 to 19.8 ± 2.5 , 89.3 ± 4.5 , 91.5 ± 5.5 and 99.8 ± 1.9 eggs/female in the control (Table 3).

Under store conditions, the percentage of eggs laid/female were significantly decreased to 11.0 ± 1.7 eggs/female after *C. chinensis* treated with *BT IP thurizide* after 120 days as compared to 98.9 ± 1.9 eggs/female in the control. In all bacterial treatments the percentage of emergence were significantly decreased to 1, 1, 1, 4 and 6 with treated with *BT IP thurizide* (Table 4).

Fig 1 show that the percentage of *C. maculatus* and *C. chinensis* significantly decreased after the end of storage and the bacteria give good protections to the cowpea seeds under store conditions.

4, Discussion

The same results obtained by [23] reported that under laboratory conditions results showed that the LC₅₀ of *Phyllotreta cruciferae*, *Pegomyia hyoscami* and *Cassidavittata* of the tested fungi *Verticillium lecanii* (V.l), *Nomuraea rileyii* (N.r) and *Paecilomyces farinosus* (P.f), respectively against the three pests ranged between 5.4×10^6 and 1.43×10^7 spores/ml. Satisfactory results with the entomopathogenic fungi were reported by [31] and [32] [33], [8] as they found that the fungi; *B. bassiana* and *M. anisopliae* reduced the LC₅₀ of *S. littoralis* under laboratory conditions.

The obtained results are similar to other studies carried out by [33] [22] and on their work on *C. Capitata* and increased the yield. These results agree with [8], [14], and [35], who proved that the application with bioinsecticides increased the yield and decreased the infestation with insect pests. Also, results were in accordance with [30] who reported that the virulence of *B. bassiana* against *C. capitata* ranged between 8 to 30% and decrease the infestation among the olive fruits. [36] recorded that *C. capitata* mortality ranged between 69 and 78% after bioinsecticides treatments.

kg/Feddan during seasons 2011 and 2012, respectively Table 4. The same results [41] control the potato tuber moth by two entomopathogenic

Paecilomyces sp. [43], [44].[42], controlled cereal aphids with entomopathogenic fungi. They found that the infestation was reduced after fungi applications under laboratory and field conditions [39], [10] , [40]and [36] found that the fungi *B. bassiana*, *M. anisopliae*, *Pacilomyces fumosoroseus* *Verticillium lecanii*; reduced insect infestations of cabbage and tomato pests under laboratory and field conditions.

The same findings obtained by ([39], [10] ,[38] , [39] [40] [44]and [13], found that the fungi *B. bassiana*, *M. anisopliae*, *Pacilomyces fumosoroseus* *Verticillium lecanii*; reduced insect infestations of cabbage and tomato pests under laboratory and field conditions. [6] found that,in all treatments the number of corn pests were significantly decreased. loss of the yield by [8] and 15), proved that applications with bioinsecticides increased the yield and decreased the infestations. Sabbour & Sahab ([39], [10] and [36]) found that the fungi reduced insect infestations of cabbage and tomato pests under laboratory and field conditions. These results agree with , ([8] and [35]), proved that applications with bioinsecticides increased the yield and decreased the the infestation with insect pests [45] and [46].

Acknowledgements.This research was supported by Agric. Department, National Research Centre, Cairo, Egypt. Project No (10120601).

Table 1. Effect of the entomopathogenic Bacteria against *Callosobruchus maculatus* larvae under laboratory conditions.

Insects	LC ₅₀ Ug/ml	Slope	Variance	95%confidence limits
<i>B.T J</i>	248	0.1	1.01	391-144
<i>B.t 0900</i>	260	0.2	1.00	214-101
<i>Bt NRRL 2172</i>	144	0.1	1.03	237-97
<i>BT IP thurizide</i>	88	0.4	0.1	129-71
<i>Bt HD112</i>	110	0.5	1.2	130-86

Table 2. Effect of the entomopathogenic Bacteria against *Callosobruchus chinensis* larvae under laboratory conditions.

Insects	LC ₅₀ Ug/ml	Slope	Variance	95%confidence limits
<i>B.T J</i>	233	0.1	1.01	369-160
<i>B.t 0900</i>	132	0.2	1.00	216-110
<i>Bt NRRL 2172</i>	145	0.1	1.03	231-93
<i>BT IP thurizide</i>	77	0.4	0.1	131-57
<i>Bt HD112</i>	100	0.5	1.2	100-81

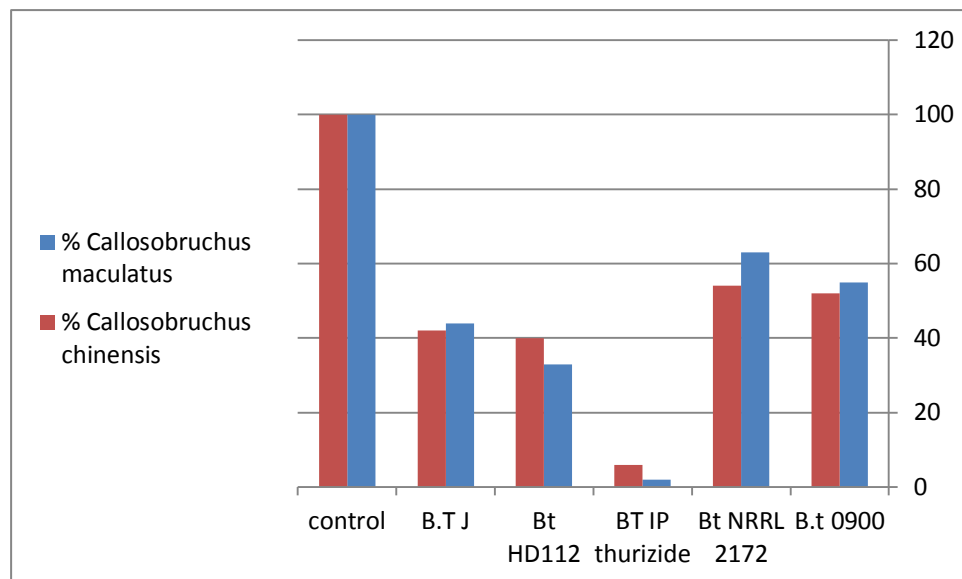
Table 3. Effect of different treatments of Bacteria on the *Callosobruchus maculatus* pests under store conditions.

Storage interval days	Control	<i>B.t 0900</i>		<i>Bt NRRL 2172</i>		<i>BT IP thurizide</i>		<i>Bt HD112</i>		<i>B.T J</i>		
	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% adult emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% adult emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)
20	19.8±2.5	87	8.0±0.5	11	11.1±2.3	10	1.7±2.7	1	3.1±7.5	1	5.5±7.0	4
45	89.3±4.5	90	25.8±8.1	20	23.8±3.8	13	5.3±0.5	1	6.0±1.9	3	7.8±2.5	10
90	91.5±5.5	95	30.8±1.5	21	35.8±7.4	27	10.2±2.3	5	11.1±4.2	10	11.0±6.5	13
120	99.8±1.9	100	36.8±2.9	32	49.8±4.5	39	11.4±3.7	7	19.0±2.8	11	21.3±0.7	21
F value	22.1		23.5		12.1		8.1		10.1		11.0	
Lsd5%	13		16		10		8		8		7	

Table 4. Effect of different treatments of bacteria *Callosobruchus chinensis* under store conditions.

Storage interval days	Control	<i>B.t 0900</i>		<i>Bt NRRL 2172</i>		<i>BT IP thurizide</i>		<i>Bt HD112</i>		<i>B.T J</i>		
	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)	no. of eggs /♀±S.E.	% emergence (F1)
20	89.9±9.9	85	8.9±9.5	11	11.0±3.5	12	1.5±1.7	1	3.3±1.5	2	5.9±7.8	5
45	92.3±4.9	91	29.8±7.3	24	24.8±6.8	16	5.2±3.5	1	6.3±0.7	4	7.9 ±8.5	11
90	94.5±5.5	96	30.8±7.5	36	39.8±7.1	28	10.0±6.5	4	11.0±2.5	11	11.8±6.8	14
120	98.9±1.9	100	39.9±1.9	39	49.8±1.5	38	11.0±1.7	6	19.0±3.5	13	21.9±8.7	22
F value	20.2		21.1		12.1		8.6		10.6		11.2	
Lsd5%	15		17		10		8		7		6	

Fig 1. Infestation percentages under store conditions of the target insect pests



REFERENCES

- [1] Songa, J.M. and W. Rono, 1998. Indigenous methods for bruchid beetle (Coleoptera: Bruchidae) control in stored beans (*Phaseolus vulgaris* L.). *Inter. J. Pest Mgt.*, 44: 1-4.
- [2] Schmale, I., F.L. Wackers, C. Cardona and S. Dorn, 2002. Field Infestation of *Phaseolus vulgaris* by *Acanthoscelides obtectus* (Coleoptera: Bruchidae), Parasitoid Abundance and Consequences for Storage Pest Control. *Environ. Entomol.*, 31: 859-863
- [3] Baby, J.K., 1994. Repellent and phagodeterrent activity of *Sphaeranthus indicus* extract against *Callosobruchus chinensis*. *Proceeding of the 6th International Working Conf. On Stored-Product protection*, 12: 17-23.
- [4] Mulungu, L.S., E.L. Luwondo, S.O.W.M. Reuben and R.N. Misangu, 2007. Effectiveness of local botanicals as protectants of stored beans (*Phaseolus vulgaris* L.) against bean bruchid (*Zabrotes subfasciatus* Boh.). *J. Entomol.*, 4: 210-217.
- [5] Nahdy, M.S. and A. Agona, 1995. Studies on the control of bean bruchids *Acanthoscelides obtectus* (Say) and *Zabrotes subfasciatus* (Bohemian) (Coleoptera: Bruchidae) in the east Africa region. In: Habtu Assefa (ed.). *Proceedings of the 25th Anniversary of Nazareth Research Center: 25 years of experience in lowland crops research*, 20-33, September, 1995. Nazareth agriculture research center, pp: 181-194.
- [6] Sabbour M.M. and abdel-Rahman. 2013. Efficacy of isolated *Nomuraea rileyi* and Spinosad against corn pests under laboratory and field conditions in Egypt *Annual review and research in biology*, 3(4): 903-912.
- [7] Sabbour, M.M., M. Ragei and A. Abd-El Rahman, 2011. Effect of Some Ecological Factors on The Growth of *Beauveria bassiana* and *Paecilomyces fumosoroseus* Against Corn Borers. *Australian Journal of Basic and Applied Sciences*, 5(11): 228-235, 2011

- [8] Sabbour, M.M. and Shadia E. Abed El-Aziz. Efficacy of some botanical oils formulated with microbial agents against the cotton leafworm and greasy cutworm attaching cotton plants. *Bull. ENT.Soc. Egypt.* 2002: 5(28): 135-151.
- [9] Sahab , A.F; Sabbour , M.M., Attallah,A.G. and Abou-Serreh, Nivin. 2014. Genetic analysis of the entomopathogenic fungus *Beauveria bassiana* to the corn borers tested by UV as physical mutagen. *International Journal of ChemTech Research* Vol.6, No.5, pp 3228-3236.
- [10] Sabbour, M.M. and Sahab, A.F. Efficacy of some microbial control agents against *Agrotis ipsilon* and *Heliothis armigera* in Egypt. *Bull. N.R.C. Egypt.* 2007: 13(33): 165-174
- [11] Alves SB. Entomopathogenic fungi. In S.B. Alves (ed.) *Microbial Control inset Fundac~ao de Estudos Luiz de Queiroz Agricultural (FEALQ) Piracicaba, Sao Paulo Brazil.* 1998;289-370 Abbott, W.W. A method of computing the effectiveness of an insecticide. *J. Econ. Entomol*1925: 18(2) (2): 265-267.
- [12] Quintela ED, McCoy C W. Effect of Imidacloprid on development locomotory response and survival of first instars of *Diaprepes abbreviatus*. *J Econ Ent.* 1997;90:988-995.
- [13]Sabbour, M.M. 2014. Evaluating Toxicity of Extracted Destruxin from *Metarhizium anisopliae* Against the grasshopper *Hetiracris littoralis* in Egypt
- [14] Sabbour M.M.2014. Biocontrol of the Tomato Pinworm *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) in Egypt. *Middle East Journal of Agriculture Research*, 3(3): 499-503.
- [15] Sabbour, M.M. 2014. Efficacy of some microbial control agents and inorganic insecticides against red flour beetle *Tribolium castaneum* and confused flour beetle, *Tribolium confusum* (Coleoptera: Tenebrionidae) Integrated Protection of Stored Products. *IOBC-WPRS Bulletin* Vol. 98, 2014.pp. 193-201.
- [16] Sabbour M.M. 2013.Efficacy of *Nomuraea rileyi* and Spinosad against olive pests under laboratory and field conditions in Egypt.*Global Journal Of Biodiversity Science And Management*, 3(2): 228-232, 2013.
- [17] Sabbour, M.M., A.A. Abd-El-Rahman and M.A. Ragei. 2013. Determinations of some extracted oils in controlling two stored product insect pests. *Middle East Journal of Agriculture Research*, Middle East Journal of Agriculture Research, 2(4): 127-132, 2013
- [18] Sabbour M.M. and M.A. AbdEl-Raheem. 2013. Repellent Effects of *Jatropha curcas*, canola and Jojoba Seed oil, against *Callosobruchus maculatus* (F.) and *Callosobruchus chinensis* (L.). *Journal of Applied Sciences Research*, 9(8): 4678-4682, 2013
- [19] Sabbour, M.M and and M.A. Abdel-Raheem. 2014. Evaluations of *Isaria fumosorosea* isolates against the Red Palm Weevil *Rhynchophorus ferrugineus* under laboratory and field conditions.*Current Science International.* 2077-4435.
- [20] Abdel-Rahman, M. A., A. Ahmed, Y. Abdel-Mallek and G.A. Hamam 2006. Comparative abundance of entomopathogenic fungi of cereal aphids in Assiut Egypt. *J. Boil. Pest Cont.;* 16(2): 39-43.

- [21] Sabbour, M.M. and Nayera, Y. Soliman. 2014. Preliminary Investigations Into The Biological Control Of Red Palm Weevil *Rhynchophorus ferrugineus* By Using three isolates of the fungus *Lecanicillium (Verticillium) lecanii* In Egypt **Volume 3 Issue 8, August 2014. 2319-7064**
- [22] Sabbour, M.M and Nayera, Y. Soliman, 2014. Evaluations of three *Bacillus thuringiensis* against *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) in Egypt. **Volume 3 Issue 8, August 2014. 2319-7064**
- [23] Sabbour, M. Mand Abdel-Rahman, A . 2007. Evaluations of some terpenes and entomopathogenic fungi on three sugar beet insect pests. J. Boil. Pest. Cont. 17:22-29
- [24]. Benz G. Environment, p. 177-214. In R. Fuxa and Y. Tanada (eds), Epizootiology of insect disease. New York, Wiley. 1987;960. Abdel-Rahman, M.A.A. Seasonal prevalence of entomo-pathogenic fungi attacking cereal aphids infesting wheat in southern Egypt. Inter. Symposium. Agric. Agadir-Morocco. 2001: 7 (10): 381-389.
- [25] Furlong MJ, Pell JK. The influence of environmental factors on the persistence of *Zoophthoradicans* conidia. J Invertebrate Path 1997; 69: 223-233.
- [26] Abdel-Rahman, M.A.A and Abdel-Mallek, A.Y... Paramilitary records on entomopathogenic fungi attacking cereal aphids infesting wheat plants in southern Egypt. First Conference for safe Alternatives to pesticides for pest managements, Assiut. 2001(12): 183-190.
- [27] Abdel-Rahman, M.A.A.; Abdel-Mallek, A.Y.; Omar S.A. and Hamam, A.H... Natural occurrence of entomopathogenic fungi on cereal aphids at Assiut. A comparison study between field and laboratory observations. Egypt. J. Boil. Sci., 2004;14(2): 107-112.
- [28] Anderson TE, Roberts DW. Compatibility of *Beauveria bassiana* strain with formulations used in Colorado potato beetle (Coleoptera: Chrysomelidae) control. Journal of Economic Entomology. 1983;76:1437-1441.
- [29] Rice RE. Bionomics of the olive fruit fly *Bactrocera (Dacus) oleae*. Univ of California Plant Prot Quart. 2000;4(2):10:-1-5.
- [30] Castillo, M.A.; Moya, P.; Hernandez, E. and Primo-Yufera, E. Susceptibility of *Ceratitidis capitata* Wiedenmann (Diptera: Tephritidae) to entomopathogenic fungi and their extract. Biol. Cont. 2000: 19(2): 274-282.
- [31] Sharaf El-Din A. Control of *Macrotoma palmate* F. adults (Coleoptera: erambycidae) by the entomogenous fungus *Beauveria bassiana* Vuill. (Deutromycotina: Hyphomycetes). Bull Ent Soc Egypt Econ Ser. 1999;26:95-107.
- [32] Sabbour., M.M. and Ismail. A. Ismail 2002. The combined effect of some microbial control agents and plant extracts against potato tuber moth *Phthorimaea operculella* (Zeller). Bull. N. R. C. Egypt. 27: 459-467.
- [33] Sabbour, Magda M, 2002. Evaluation studies of some bio-control agents against corn borers in Egypt. Annal Agric. Sci. Ain Shams Univ. Cairo, 47(3): 1033-1043

[34]Espin, G.A. T. laghi De .S.M., Messias, C.L. and Pie-Drabuena, A.E. Pathogenicidad de *Metarhizium anisopliae* diferentes fases de desenvolvimen to de *Ceratitis capitata*(Wied.) (Diptera: Tephritidae). Revista Brasileria de Entomologia. 1989 :33(2): 17-23.

[35] Sabbour, M.M. and Shadia, E. Abd-El-Aziz. Efficacy of some bioinsecticides against *Bruchidius incarnatus*(BOH.) (Coleoptera: Bruchidae) Infestation during storage. J. Plant Prot. Res. 2010: 50 (1): 28-34.

[36]Sahab, A.F. and Sabbour, M.M. Virulence of four entomo-pathogenic fungi on some cotton pests with especial reference to impact of some pesticides, nutritional and environmental factors on fungal growth. Egypt. J. Boil. Pest Cont. 2011: 21 (1): 61-67.

[37]Abdel-Rahman MAA, Abdel-Mallek AY. Paramilitary records on entomopathogenic fungi attacking cereal aphids infesting wheat plants in southern Egypt.First Conference for safe Alternatives to pesticides for pest managements, Assiut.2001;(12):183-190.

[38]Sabbour, M. M.2007.Effect of some natural bioagents and natural enemies against aphids in wheat fields J. Boil. Pest.Cont 33: 33-39.

[39]Sabbour, M.M. and Sahab, A.F. Efficacy of some microbial control agents against cabbage pests in Egypt. Pak. J. Biol. Sci. 2005:5(8): 1351-1356.

[40] Tanda, Y. and Kaya, H.K... Insect Pathology.Academic Press, San Diego, CA, USA. 1993: 3(4): 123-129.

[41] Sabbour M.M. and Singer, S.M.2014. Evaluations of two isolated *Paecilomyces*against *Phthorimaea operculella* (Lepidoptera: Gelechiidae) under laboratory and field conditions Volume 3 Issue 9, september 2014. 319-324

[42]Abdel-Rahman MAA, Abdel-Mallek AY, Omar SA, Hamam AH. Natural occurrence of entomopathogenic fungi on cereal aphids at Assiut.A comparison study between field and laboratory observations. Egypt J Boil Sci. 2004;14(2):107-112.

[43]Abdel-Rahman MAA. Seasonal prevalence of entomo-pathogenic fungi attacking cereal aphids infesting wheat in southern Egypt. Inter Symposium Agric Agadir- Morocco. 2001;7(10):381-389.

[44] Sabbour M.M. and Singer, S.M. 2014. Efficacy of Two Entomopathogenic Fungi against Corn Pests Under Laboratory and Field Conditions in Egypt. European Journal of Academic Essays 1(9): 1-6, 2014.

[45] Nichimbi-Msolla, S. and R.N. Misangu, 2002. Seasonal distribution of common bean bruchid species in selected areas in Tanzania. Proceedings of the Bean Seed Workshop, Arusha, Tanzania, 12–14 January, 2001. Bean/Cowpea Collaborative Research Support Program–East Africa. pp. 5.

[46] Emanu, Getu, Ahmed Ibrahim and Fridissa Iticha, 2003. Review of lowland pulse insect pest research in Ethiopia. Proceedings of grain legume workshop, 22-27 September, 2003, Addis Ababa, Ethiopia.

IJSER